

# Some Factors Affecting the Efficiency of Hybrid Embryo Rescue in the ‘Shiranuhi’ Mandarin

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## Abstract

The ‘Shiranuhi’ mandarin has attractive characteristics such as a high soluble solid content, a lack of seeds, and a rind that is easy to peel. Thus, it has been utilized as a genetic resource in citrus breeding programs. However, it has a polyembryony trait that causes problems when producing sexual hybrids. Therefore, methods need to be established to distinguish between zygotic and nucellar plants from cross offspring. In this study, the ‘Shiranuhi’ mandarin and the ‘Sanguinelli’ blood orange were crossed to produce immature embryos that were used to detect zygotic embryos at 90, 105, 125, 145, and 180 days after pollination (DAP). For germination, embryos at different developmental stages were cultured on Murashige and Skoog (MS), Murashige and Tucker (MT), and Gamborg B5 media supplemented with 500 mg·L<sup>-1</sup> malt extract, 25 mg·L<sup>-1</sup> adenine sulfate (ADS), 1 mg·L<sup>-1</sup> gibberellic acid (GA<sub>3</sub>), 3% sucrose (w/v), and 0.8% phyto agar (w/v). The germination rate of immature embryos ranged from 36.4% to 74.9% depending on DAP, while germination rate was not significantly different by medium type. The rooting rate was high during the late embryo developmental stages and when there were low sucrose concentrations and high GA<sub>3</sub> concentrations. The percentage of zygotic plants per seed was measured using simple sequence repeat (SSR) markers and averaged 12.1% for 90 DAP, 7.6% for 105 DAP, 6.8% for 125 DAP, 1.0% for 145 DAP, and 4.1% for 180 DAP. These results indicated that embryo developmental stage was the greatest factor in relation to inorganic salt, sucrose, and GA<sub>3</sub> concentration for increasing the selection efficiency of sexual hybrids in crosses using polyembryony maternal parents by embryo rescue and SSR marker analysis.

**Additional key words:** embryo culture, polyembryony, sexual hybrid, simple sequence repeats, zygotic plant

## Introduction

Korean citrus production predominantly comprises satsuma mandarins (*Citrus unshiu*), which account for approximately 90% of total production (JSGP, 2016). The total production area of citrus

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has declined by approximately 28% from 26,234 ha in 2002 to 20,523 ha in 2015. The production of late maturing varieties reached 67,406 MT for 2,112 ha in 2015, an increase of 10% in the past 10 years. High yield production of satsuma mandarins combined with alternate year bearing and poor quality has repeatedly resulted in overproduction. This has led to price instability and cultivation decline. On the contrary, fruit imports in Korea have steadily increased due to the implementation of many Free Trade Agreements since 2003. Two major fruit imports were orange and banana, which have been popular with Korean consumers due to price competitiveness and good flavor. Therefore, it is urgent and essential for the Korean citrus industry to develop new, high-quality varieties with a high amount of bioactive components and a high tolerance to biotic and abiotic stresses to stay competitive in the market and satisfy not only consumer needs but also grower demand.

Historically, new varieties have mostly depended on the selection of nucellar seedlings (apomixis) and bud sports. The long juvenility from seed to flowering, complicated gene composition, polyembryony, gamete sterility, and self-incompatibility are significant barriers to developing new promising hybrid varieties in citrus breeding (Soost and Cameron, 1975). Therefore, it is necessary to develop methods to distinguish between zygotic and nucellar plants after sexual cross. Identification of sexual hybrids obtained from the cross using polyembryony maternal parents is difficult even though parental lines have convenient dominant traits (Golein et al., 2011). Although morphological (Cameron, 1979), infrared spectroscopy (Pieringer and Edwards, 1967), isozymatic (Anderson et al., 1991), chromatographic (Tatum et al., 1974), and molecular marker approaches such as restriction fragment length polymorphism (RFLP) (Carimi et al., 1998) and random amplified polymorphic DNA (RAPD) (Rodriguez et al., 2005; Yun et al., 2007; Mondal and Saha 2013) can be used to distinguish zygotic and nucellar seedlings, these methods are limited by low reproducibility or efficiency. Hence, advanced molecular markers such as simple sequence repeats (SSRs) (Oliveira et al., 2002; Tan et al., 2007; Yildiz et al., 2013) and inter-simple sequence repeat (ISSRs) (Golein et al., 2011) have been developed.

Embryo culture is one of the earliest forms of in vitro culture (Bridgen, 1994). Since the first attempt by Ohta and Furusato (1957), immature embryo rescue of *Citrus* has been useful in producing sexual hybrids through distinguishing zygotic and nucellar plants in many polyembryony types of *Citrus* (Carimi et al., 1998; Pérez-Tornero and Porras, 2008). In the Korean citrus breeding program, many polyembryony cultivars such as satuma, ponkan, and 'Shiranuhi' mandarins have also been widely utilized. These mandarins are known to have higher soluble solid contents, seedlessness, loose skin, or sweet taste (Matsumoto, 2001) and are attractive as breeding materials, except for polyembryony. Using RAPD analysis, Yun (2007) reported that the 'Shiranuhi' mandarin has a low frequency of sexual hybrids of 0.0 to 3.9%, which was lower than that of the satuma mandarin at 8.6 to 13.4%.

In this study, embryo rescue stage, culture medium, and sucrose and GA<sub>3</sub> concentrations in the embryo culture were evaluated with the aim to improve the efficiency of obtaining sexual hybrids in 'Shiranuhi' mandarins with a polyembryony trait.

## Materials and Methods

### Plant Materials and Preparation of Embryo Rescue

Artificial pollination was performed at Hannong Bio Industry (HBI) Breeding Orchard in 2014 to 2016. The 'Shiranuhi' mandarin [*(C. unshiu × C. sinensis) × C. reticulata*] at full bloom stage was used as the maternal parent and the

‘Sanguinelli’ blood orange (*C. sinensis*) was used as the paternal parent. Immature fruits of ‘Shiranuhi’ were harvested at 90, 105, 125, 145, and 180 days after pollination (DAP). After washing fruits carefully with tap water and blot-drying, fruits were immersed in 70% (v/v) ethanol for 30 min, rinsed three times with sterile distilled water, and dried on a clean bench. The fruits were cut open, and immature seeds were taken out by forceps. Immature embryos were separated carefully from the microphyll end of the seeds under a light microscope.

### Culture Media and Sucrose and GA<sub>3</sub> Concentration Supplementation

MS (Murashige and Skoog, 1962), MT (Murashige and Tucker, 1969), and B5 (Gamborg et al., 1968) media were used to investigate the effect of medium type on germination and rooting of embryos. Each medium contained 500 mg·L<sup>-1</sup> malt extract (ME), 25 mg·L<sup>-1</sup> adenine sulfate (ADS), 1 mg·L<sup>-1</sup> GA<sub>3</sub>, 3% sucrose (w/v), and 0.8% phyto agar (w/v). The pH of the medium was adjusted to 5.8 before autoclaving (121°C for 15 min), and GA<sub>3</sub> was added to the medium through filtering after autoclaving. The embryo culture was maintained at 25 ± 2°C under a 16-h-light/8-h-dark photoperiod. After 2 to 3 weeks, the germination and rooting rate of the embryos were measured. After that, the germinated embryos were transferred to MS medium supplemented with 1 mg·L<sup>-1</sup> benzyl adenine (BA), 0.1 mg·L<sup>-1</sup> naphthalene acetic acid (NAA), 3% sucrose (w/v), and 0.8% phyto agar (w/v) at pH 5.8.

Different concentrations of GA<sub>3</sub> and sucrose in the MS medium were also evaluated. Embryos at 90 DAP were placed on the MS media supplemented with sucrose (3, 5, 7, 9, and 12%) and GA<sub>3</sub> (0, 0.5, 1.0, 2.5, and 5.0 mg·L<sup>-1</sup>). The survival, germination, and rooting rate of the embryos were measured 4 weeks after the embryo culture. After that, the germinated embryos were transferred to MS medium supplemented with 1 mg·L<sup>-1</sup> BA, 0.1 mg·L<sup>-1</sup> NAA, 3% sucrose (w/v), and 0.8% (w/v) phyto agar (w/v) at pH 5.8.

### Genomic DNA Extraction and SSR Marker Analysis

Genomic DNA was extracted from young leaves of the embryo-rescued plants using the Biomedic<sup>®</sup> gDNA Extraction Kit (Biomedic Co., Ltd., Korea). Genotyping was performed using the M13-tailed PCR method (Schuelke, 2000). Three SSR markers showing polymorphism between ‘Shiranuhi’ and ‘Sanguinelli’ were obtained from a previous report (Woo et al., 2019). The primers to amplify three SSR loci used in this study were as follows: BM-CiSSR94 forward (5'-TGTA AACGACGGCCAGTGAATTGGGAGG ACGAACTGA-3') and reverse (5'-CGAGCCCTAGACAGAGATGG-3'), BM-CiSSR 111 forward (5'-TGTA AACGACGGCCAGTCCGATACAGCACAAAGCAAA-3') and reverse (5'-TGGAAAGAGA GAAGCCAAGC-3'), and BM-CiSSR115 forward (5'-TGTA AACGACGGCCAGTCGGTGTGTATTGGGTACACG-3') and reverse (5'-TGGAAAGAGAGAAGCCAAGC-3'). PCR amplification was conducted using the ABI 2720 thermal cycler (Applied Biosystems) in a total volume of 10 mL containing 20 ng DNA, 5 mL 2x HS<sup>TM</sup> Taq mix (containing 0.3 unit/mL HS Taq DNA polymerase, 3.2 mM MgSO<sub>4</sub>, and 0.4 mM dNTPs) (Dongsheng Biotech, Guangzhou, China), each 0.2 mL of 10 pmol M13-tailed forward primer, 1 mL of 10 pmol reverse primer, and 1 mL of 10 pmol 6-FAM labeled M13 primer (5'-6-FAM-TGTA AACGACGGCCAGT-3'). The conditions for PCR amplification were as follows: 5 min for initial denaturation at 94°C; 15 cycles of 30 s at 94°C, 30 s at 58°C, and 30 s at 72°C; followed by 20 cycles of 30 s at 94°C, 30 s at 53°C, and 30 s at 72°C; concluding with 1 cycle of 30 min at 72°C as a final extension reaction. The amplified fragments were separated by capillary electrophoresis on an ABI 3730 DNA analyzer (Applied Biosystems) using a 50-cm capillary with a DS-33 install standard as a matrix. Analysis of allele sizes was conducted using GeneMapper

software (ver. 4.0; Applied Biosystems).

### Statistical Analysis

The obtained data were analyzed using R console 3.2.3, and significant differences between means were assessed using Duncan’s multiple range tests at  $p \leq 0.05$ .

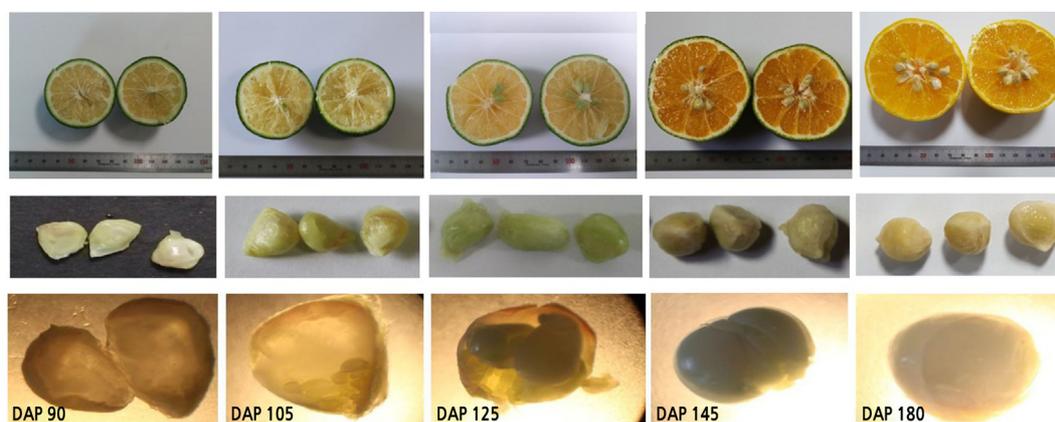
## Results and Discussion

### Fruit Set, Seed Formation, and Embryo Development

Fruit sets of the ‘Shiranuhi’ mandarin cross-pollinated with the ‘Sanguinelli’ blood orange for 2014–2016 were recorded with an average of 44.5% after a physiological drop, which had yearly variation depending on weather conditions (Table 1). Several studies reported that the fruit set of the cross-pollinated citrus might be affected by crossing parent and environmental conditions such as rootstock and climatic conditions (Soost and Cameron, 1975; Kedar and Gopal, 1977; Yelenosky, 1985; Sharma et al., 1999). The number of normal seeds per fruit ranged from 2.9 to 7.5 for 3 years with some variations. The number of normal seeds per matured fruit observed in this study was higher than those previously reported by Yun (2007), who obtained an average of 0.5 to 1.3 full seeds in ‘Shiranuhi, pollinated with ponkan mandarin and ‘Swingle’ citrumelo. This might have been caused by the difference in pollen genotype, but the exact reason requires further study.

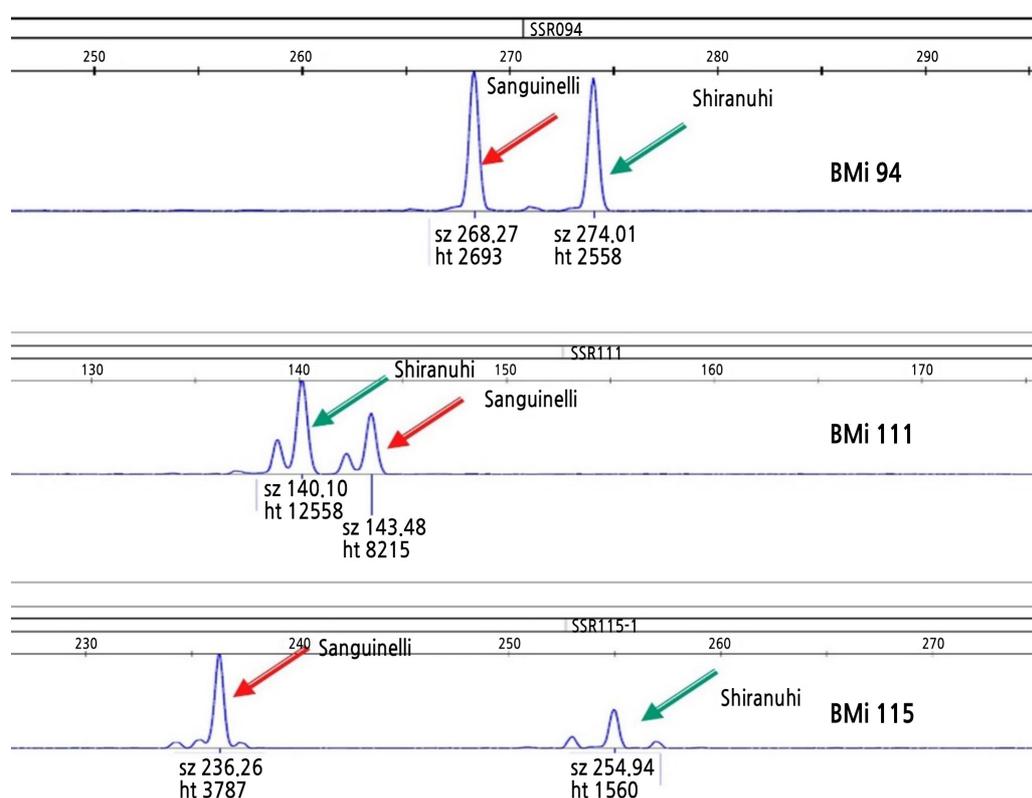
**Table 1.** Fruit set and seed formation in the ‘Shiranuhi’ mandarin [(*Citrus unshiu* × *C. sinensis*) × *C. reticulata*] pollinated with ‘Sanguinelli’ blood orange (*C. sinensis*)

Year	No. of crosses	No. of fruits	Fruit set (%)	Total no. of seeds	No. of seeds per fruit
2014	250	135	54.0	397	2.9
2015	668	314	47.0	2,346	7.5
2016	342	111	32.5	795	7.2
Avg.	420	186.7	44.5	1,179.3	5.9



**Fig. 1.** Morphological development of fruits, seeds, and embryos at different days after pollination (DAP) in the ‘Shiranuhi’ mandarin [(*C. unshiu* × *C. sinensis*) × *C. reticulata*] pollinated with the ‘Sanguinelli’ (*C. sinensis*) blood orange.

The total number of seeds ranged from 8.4 to 12.7 with some variation depending on the fruit development stage and DAP; the average number of seeds was the lowest, 8.4, at 180 DAP and the highest, 12.7, at 90 DAP (Table 2). The number of normal seeds decreased with increased DAP, whereas the number of abnormal seeds, the length of the seeds, and the number of embryos per seed were significantly increased as DAP increased. Furausato et al. (1957) showed that the correlation coefficient between the number of seeds contained in the fruit and the mean embryo number per seed fluctuated irregularly from year to year, from tree to tree, and from branch to branch, and no regularity could be observed. In sour orange, none of the embryos were observed in the immature seeds harvested at 65 – 85 DAP, and the average number of dissected embryos per seed was significantly affected by developmental stage and genotype (Carimi et al.,



**Fig. 2.** Diagrams of SSR loci amplified with BM-CiSSR 94, BM-CiSSR 111, and BM-CiSSR 115 primers in the ‘Shiranuhi’ mandarin [(*C. unshiu* × *C. sinensis*) × *C. reticulata*] and the ‘Sanguinelli’ (*C. sinensis*) blood orange.

**Table 2.** Seed and embryo development at different developmental stages of the ‘Shiranuhi’ mandarin [(*C. unshiu* × *C. sinensis*) × *C. reticulata*] pollinated with the ‘Sanguinelli’ blood orange (*C. sinensis*) cross

DAP <sup>z</sup>	Total no. of seeds	No. of normal seeds	No. of abnormal seeds	Length of seed (mm)	No. of embryos per seed
90	12.7 ± 5.1 a <sup>y</sup>	12.3 ± 5.1 a	0.4 ± 0.6 c	6.1 ± 0.9 d	9.0 ± 7.4 c
105	10.4 ± 5.4 ab	10.1 ± 5.1 ab	0.3 ± 0.7 c	7.6 ± 0.9 c	9.0 ± 2.9 c
125	10.4 ± 4.8 ab	8.2 ± 4.7 bc	2.2 ± 3.3 b	8.8 ± 1.1 b	13.6 ± 5.7 b
145	11.9 ± 4.9 ab	7.2 ± 4.0 bc	4.7 ± 3.3 a	9.5 ± 1.1 ab	16.6 ± 5.3 a
180	8.4 ± 5.7 b	5.4 ± 5.0 c	3.0 ± 2.9 b	10.1 ± 1.2 a	17.2 ± 6.5 a

<sup>z</sup>DAP: days after pollination.

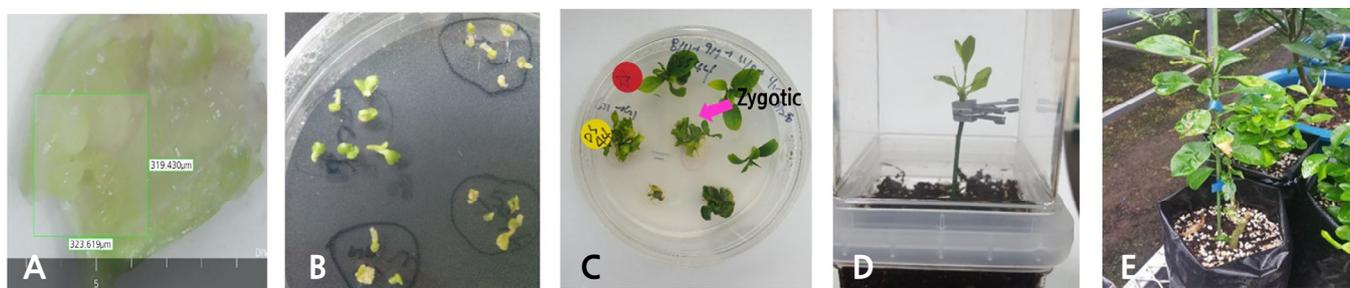
<sup>y</sup>Mean separation within column by Duncan’s multiple range test at  $p < 0.05$ .

1998). Therefore, it might be considered that the appropriate time for embryo rescue is different depending on citrus genotype.

The endosperm tissues were easily recognized by 125 DAP (Fig. 1). Peel coloration started at 180 DAP and, at that time, most seeds were in the form of a whole seed. This result indicated that there might be a correlation between coloration of the pericarp and full development of the seeds. The number of normal seeds in cross-pollinated fruits of the ‘Shiranuhi’ mandarin decreased as the fruits developed, whereas the number of abnormal seeds increased (Table 2). This phenomenon might be related to the natural seed degeneration process during fruit development. Button and Kochba (1977) indicated that polyembryony cultivars usually contain a zygotic embryo at the earlier stages of ovule and subsequent seed development. Nucellar embryos are most vigorous and so inhibit the full growth of the zygotic embryo and cause its degeneration before seed maturation.

### Embryo Germination and Rooting Response to Embryo Rescue Time and Culture Medium Type

To examine the embryo germination response of the ‘Shiranuhi’ mandarin cross-pollinated with ‘Sanguinelli’ blood orange, embryos were excised from seeds at 90–180 DAP and then were subjected to in vitro culture on MT, MS, and B5 media. The embryos developed into the cotyledon stage or shoots after 7 days of exposure to the medium (Fig. 3). The germination rate of the embryo increased as DAP increased and ranged from 22.0 to 76.1% with the lowest at 90 DAP (avg. 36.4%) and the highest at 145 DAP (avg. 74.9%) (Table 3). Rooting had not occurred in embryos obtained from 90–105 DAP, whereas it was remarkably increased from 28.3% at 125 DAP to 64.8% at 180 DAP (Table 3). As a result, there was a significant difference in germination and rooting rate depending on embryo developmental stage and embryo rescue time with DAP, but there was no difference depending on culture medium type. These results indicated that embryo germination and rooting are strictly correlated with embryo development. This result was in accordance with a previous study (Carimi et al., 1998) that reported that there was a positive effect between embryo age and embryo germination, and the frequency of embryo germination and plant formation also increased with increasing age. Bridgen (1994) also reported that precocious germination (the germination of embryos before the completion of normal embryo development) causes the establishment of weak seedlings, which was similar to our result that showed low root activity at early embryo development stages.



**Fig. 3.** Overall process of embryo rescue from an initial stage of embryo culture to acclimatization stage by ex vitro grafting in the ‘Shiranuhi’ mandarin [(*C. unshiu* × *C. sinensis*) × *C. reticulata*] pollinated with the ‘Sanguinelli’ (*C. sinensis*) blood orange. (A) Embryo excised at 90 days after pollination (DAP). (B) Embryos cultured for 2 weeks in MS media containing 500 mg·L<sup>-1</sup> malt extract (ME), 25 mg·L<sup>-1</sup> adenine sulfate (ADS), 5 mg·L<sup>-1</sup> gibberellin 3 (GA<sub>3</sub>), and 3% sucrose (w/v). (C) Embryos at the screening stage by the SSR marker analysis. (D) Ex vitro grafting of the zygotic origin plant on trifoliolate orange rootstock. (E) Zygotic origin plant grown under greenhouse conditions.

**Table 3.** Effect of embryo developmental stage and medium type on embryo germination and rooting rate after 2-3 weeks of embryo culture in the 'Shiranuhi' mandarin [(*C. unshiu* × *C. sinensis*) × *C. reticulata*] pollinated with the 'Sanguinelli' blood orange (*C. sinensis*)

Days after pollination (DAP)	Medium	Germination (%)	Rooting (%)
90	MT	45.0 ± 24.5 <sup>z</sup>	0
	MS	42.1 ± 19.6	0
	B5	22.0 ± 16.8	0
105	MT	58.0 ± 3.8	0
	MS	61.8 ± 11.6	0
	B5	54.4 ± 15.3	0
125	MT	69.6 ± 8.8	28.3 ± 12.1
	MS	73.3 ± 11.5	29.7 ± 14.0
	B5	66.0 ± 8.7	28.6 ± 12.6
145	MT	74.6 ± 7.9	48.5 ± 7.4
	MS	73.9 ± 5.6	61.2 ± 3.9
	B5	76.1 ± 10.9	56.3 ± 8.1
180	MT	66.9 ± 6.6	55.0 ± 7.3
	MS	72.3 ± 10.9	58.1 ± 11.9
	B5	72.9 ± 6.3	64.8 ± 6.4
DAP		***	***
Significance Medium		NS	NS
DAP × Medium		NS	NS

<sup>z</sup>Values represent mean ± SE (n = 15).

NS,\*\*\*,\*\*\*\* Not significant or significant at 0.05, 0.01, or 0.001, respectively.

### Embryo Germination and Rooting Response to Sucrose and GA<sub>3</sub> Concentration

The essential role of GA on seed development can be inferred from transgenic and mutant studies in *Arabidopsis*, tomato, and pea, where depletion of bioactive GA from seed tissues caused abortion at early stages of development (Plackett and Wilson, 2016). Several studies have also reported that GA<sub>3</sub> and sucrose were more useful for embryo rescue of *Citrus* (Carimi et al., 1998; Usman et al., 2002). In this study, it was investigated whether 'Shiranuhi' mandarin cross-pollinated with 'Sanguinelli' blood orange showed a similar response to sucrose and GA<sub>3</sub> concentration (Table 4). The highest survival rate was 99.7%, and this was obtained from the combination of 12% sucrose and no supplement with GA<sub>3</sub>. The highest germination rate was 79.0%, and this was achieved with 5% sucrose and 5 mg·L<sup>-1</sup> GA<sub>3</sub>. Survival and germination rate of embryos varied with combinations of sucrose and GA<sub>3</sub> concentration, which showed a complicated response with no constant tendency. Rooting rate increased as sucrose concentration decreased while GA<sub>3</sub> concentration was high (Table 4). The highest rooting rate 21.5% was obtained with the combination of 3% sucrose and 5 mg·L<sup>-1</sup> GA<sub>3</sub>.

Bridgen (1994) indicated that mature embryos usually grow well in medium with 2 - 3% sucrose, whereas immature embryos grow better with a higher concentration of 8 - 12%, which mimics the high osmotic potential within the young embryo sac. Usman et al. (2002) also reported that the germination rate of embryos in an interploidy hybridization of 'Kinnow' mandarin and 'Succari' orange was affected by GA<sub>3</sub> concentration. GA<sub>3</sub> plays various roles either promoting or inhibiting shoot and root formation depending on the species or conjunction with the auxin (Moshkovy, 2008). Eshed et al.

(1996) reported that pretreatment of bark of a 3-year-old stock oak tree (*Quercus ithaburensis*) with GA<sub>3</sub> increased rooting over the control by 6- to 7-fold. Also, Ford et al. (2002) reported that the induction of adventitious rooting in cuttings of cherry (*Prunus avium*) was stimulated by GA<sub>3</sub> pretreatment and that the number of roots per rooted cutting was increased up to 80% or more. Sagee et al. (1990) indicated a great enhancement of rooting in citrus by GA treatment in cuttings. Gibberellins are generally associated with juvenility, and application of gibberellins to the mature phase of several species might induce some juvenile characteristic (Hackett, 1985). Cuttings and tissue culture are vegetative propagation, and the response of immature rescued embryos to GA treatment might be similar to the response of cuttings. In this study, the embryo rescue response to sucrose and GA<sub>3</sub> concentration was evaluated only at the young developmental stage from one genotype. Therefore, further study is required to determine how its response might be different with other genotypes and embryo developmental stages.

**Table 4.** Effect of sucrose and GA<sub>3</sub> concentration on the survival, germination, and rooting rate of embryos obtained at 90 DAP from the 'Shiranuhi' mandarin [(*C. unshiu* × *C. sinensis*) × *C. reticulata*] pollinated with the 'Sanguinelli' blood orange (*C. sinensis*) after 4 weeks of embryo culture

Sucrose (g·L <sup>-1</sup> )	GA <sub>3</sub> (mg·L <sup>-1</sup> )	Survival (%)	Germination (%)	Rooting (%)
30	0	73.0 ± 9.1 <sup>z</sup>	23.6 ± 15.6	0
	0.5	72.8 ± 24.8	39.3 ± 20.9	4.2 ± 4.3
	1	87.8 ± 5.9	60.7 ± 8.4	5.0 ± 1.7
	2.5	70.9 ± 10.3	54.7 ± 10.8	3.8 ± 5.4
	5	69.6 ± 6.1	59.0 ± 4.3	21.5 ± 13.0
50	0	91.2 ± 8.3	41.5 ± 20.2	0
	0.5	79.8 ± 3.3	40.1 ± 13.0	1.1 ± 0.9
	1	71.1 ± 20.8	50.0 ± 11.4	1.3 ± 1.8
	2.5	98.5 ± 1.4	66.4 ± 5.1	5.6 ± 2.6
	5	94.9 ± 4.2	79.0 ± 7.9	15.2 ± 3.2
70	0	97.5 ± 3.0	63.5 ± 4.6	0
	0.5	66.9 ± 5.8	49.5 ± 5.0	4.1 ± 4.3
	1	86.8 ± 14.9	54.1 ± 7.3	0
	2.5	93.7 ± 5.5	57.6 ± 13.5	5.3 ± 6.4
	5	96.5 ± 0.4	64.8 ± 6.7	2.4 ± 1.8
90	0	94.2 ± 5.0	68.1 ± 3.8	0
	0.5	99.5 ± 0.7	69.3 ± 10.4	0
	1	92.3 ± 4.4	70.3 ± 5.5	0
	2.5	98.3 ± 1.4	63.2 ± 18.1	0
	5	81.4 ± 3.5	54.3 ± 14.1	0.4 ± 0.5
120	0	99.7 ± 0.5	67.2 ± 0.9	0
	0.5	93.3 ± 9.4	57.4 ± 18.6	0
	1	90.7 ± 8.2	51.7 ± 4.5	0
	2.5	97.8 ± 3.1	66.2 ± 14.0	0
	5	97.4 ± 2.4	48.9 ± 15.7	1.3 ± 1.9
Significance	Sucrose	**	NS	*
	GA <sub>3</sub>	NS	NS	***
	Sucrose × GA <sub>3</sub>	*	*	**

<sup>z</sup>Values represent mean ± SE (n = 15).

NS,\*,\*\*,\*\* Not significant or significant at  $p \leq 0.05$ , 0.01, or 0.001, respectively.

### Frequency of Zygotic Embryo Rescue through SSR Marker Analysis

The zygotic individuals from the sexual cross between the ‘Shiranuhi’ mandarin and the ‘Sanguinelli’ blood orange were selected with SSR markers developed previously (Woo et al., 2019). Fig. 2 shows a representative SSR analysis, and the results are represented in Table 5. The zygotic embryo percentage per seed ranged from 12.1% at 90 DAP to 1.0% at 145 DAP and 4.1% at 180 DAP with a decrease as embryos developed further. This result showed that zygotic individuals could be selected at a higher rate from embryo rescue at the early stage compared to that at the late stage.

Several studies have reported various zygotic frequencies in other citrus genotypes depending on the maternal and pollen genotypes, which ranged from 0 – 87% (Frost and Soost, 1967; Hwang, 1991; Bastianel et al., 1998; García et al., 1999; Ruiz et al., 2000; Yun et al., 2007; Yildiz et al., 2013; Jin et al., 2015). Andrade-Rodríguez (2004) reported that the zygotic embryos were located near the micropylar end and were the smallest mature seeds of *C. volkameriana*. In contrast, Yun (2007) showed that positioning patterns of hybrids of mature ‘Miyagawa Wase’ and ‘Okitsu Wase’ are different depending on the cultivar. In seeds of sour orange collected at 125 – 220 DAP, a zygotic embryo was mixed with many nucellar embryos, and they could not be distinguished from each other on the bases of size and position in the seed (Carimi et al., 1998). These results showed that there is a limit to the morphological selection method based on size and location of embryos cultured from mature citrus seeds.

Rangan et al. (1969) could successfully select hybrid plants from the polyembryonic sour orange *C. aurantium*. Sexual hybrid plants were obtained from the embryos at the heart-shaped stage of immature seeds at 100 – 120 days after anthesis. They also reported that varieties containing few embryos showed the formation of more seeds with monoploidy and more healthy development of zygotic embryos, further increasing the zygotic ratio. Carimi et al. (1998) also mentioned that the zygotic embryo rescue appeared to be dependent on the number of nucellar embryos. Lower numbers of nucellar embryos per seed usually results in a larger dimension and greater probability of zygotic embryo survival. Our results also showed that the number of embryos from the ‘Shiranuhi’ mandarin cross-pollinated with the ‘Sanguinelli’ blood orange was remarkably increased as DAP increased, and the higher rate of the zygotic embryo was obtained from immature embryo rescue at the earlier stage.

In conclusion, immature embryo rescue could be a preferred method for early selection of citrus hybrids. The reason for its superiority is that citrus plants take several years to bear fruit, which makes it difficult to conduct early characterization and requires a large-scale experimental field, as well as considerable time and effort to manage them. The goal of citrus

**Table 5.** Frequency of zygotic origin embryo rescued at different embryo developmental stages and identified by SSR marker analysis in the ‘Shiranuhi’ mandarin [*(C. unshiu* × *C. sinensis*) × *C. reticulata*] pollinated with the ‘Sanguinelli’ (*C. sinensis*) blood orange

DAP <sup>z</sup>	No. of seeds (plants)	Zygotic origin <sup>y</sup>	Zygotic origin per seed (plants)
90	58 (134)	7	12.1 (5.2)
105	118 (614)	9	7.6 (1.5)
125	103 (858)	7	6.8 (0.8)
145	98 (675)	1	1.0 (0.1)
180	97 (576)	4	4.1 (0.7)

<sup>z</sup>DAP: days after pollination.

<sup>y</sup>The ratio of zygotic origin was calculated as the total number of SSR genotyping detected shoot per total number of seeds or embryos evaluated × 100.

breeding in Korea is to improve the sugar content, eradicate the seed, and shorten the time to maturity, in addition to increasing the aroma and resistance to pests and disease. The *C. unshiu* fruits have valuable traits such as seedlessness, early maturity, and a rind that is easy to peel. Therefore, it would be interesting to introduce our method to *C. unshiu* and determine if it can improve selection of zygotic plants.

## Literature Cited

- Anderson CM, Castle WS, Moore GA (1991) Isozymic identification of zygotic seedlings in Swingle citrumelo *Citrus paradise* × *Poncirus trifoliata* nursery and field populations. *J Am Soc Hortic Sci* 116:322-326. doi:10.21273/JASHS.116.2.322
- Andrade-Rodríguez M, Villegas-Monter A, Carrillo-Castañeda G, García-Velázquez A (2004) Polyembryony and identification of Volkamerian lemon zygotic and nucellar seedlings using RAPD. *Pesq Agropec Bras* 39:551-559. doi:10.1590/S0100-204X2004000600006
- Bastianel M, Schwarz SF, Colleta-Filho HD, Lin LL, Machado MA, Koller OC (1998) Identification of zygotic and nucellar tangerine seedlings (*Citrus* spp.) using RAPD. *Genet Mol Biol* 21:123-127. doi:10.1590/S1415-47571998000100020
- Bridgen MP (1994) A review of plant embryo culture. *HortScience* 29:1243-1246. doi:10.21273/HORTSCI.29.11.1243
- Button J, Kochba J (1977) Tissue culture in the citrus industry. In J Reinert, YPS Bajaj, eds, *Applied and Fundamental Aspects of Plant Cell, Tissue and Organ Culture*. Springer-Verlag, pp 70-92
- Cameron JW (1979) Sexual and nucellar embryony in F1 hybrids and advanced crosses of Citrus and Poncirus. *J Am Soc Hortic Sci* 104:408-410
- Carimi F, de Pasquale F, Puglia AM (1998) In vitro rescue of zygotic embryos of sour orange, *Citrus aurantium* L., and their detection based on RFLP analysis. *Plant Breed* 117:261-266. doi:10.1111/j.1439-0523.1998.tb01936.x
- Eshed Y, Riov J, Atzmon N (1996) Rooting oak cuttings from gibberellin-treated stock plants. *HortScience* 31:872-873. doi:10.21273/HORTSCI.31.5.872
- Ford Y-Y, Taylor JN, Blake PS, Marks TR (2002) Gibberellins A<sub>3</sub> stimulates adventitious rooting of cuttings from cherry (*Prunus avium*). *Plant Growth Regul* 37:127-133. doi:10.1023/A:1020584627919
- Frost HB, Soost RK (1967) Seed reproduction: Development of gametes and embryos. In W Reuther, LD Batchelor, HJ Webber, eds, *The Citrus Industry*. University of California Press, Berkeley, CA, USA, pp 290-324
- Furusato K, Ohta Y, Ishibashi K (1957) Studies on polyembryony in *Citrus*. *Seiken Ziho, Rpt Kihara Inst Biol Res* 23:40-48. doi:10.1080/00220973.1939.11018061
- Gamborg OL, Miller RA, Ojima K (1968) Nutrient requirements of suspension cultures of soybean root cells. *Exp Cell Res* 50:151-158. doi:10.1016/0014-4827(68)90403-5
- García R, Asín MJ, Forner J, Carbonell EA (1999) Genetic analysis of apomixes in Citrus and Poncirus by molecular markers. *Theor Appl Genet* 99:511-518. doi:10.1007/s001220051264
- Golein B, Fifaei R, Ghasemi M (2011) Identification of zygotic and nucellar seedlings in *citrus* interspecific crosses by inter simple sequence repeats (ISSR) markers. *Afr J Biotechnol* 10:18965-18970
- Hackett WP (1985) Juvenility, maturation and rejuvenation in woody plants. *Hortic Rev* 7:109-155. doi:10.1002/9781118060735.ch3
- Hwang AS (1991) The polyembryony and identification of zygotic seedlings of lemon. *J Agric Res China* 40:225-232
- Jeju Special Self-Governing Province (JSGP) (2016) Citrus annual distribution processing analysis. Jeju special self-governing province citrus marketing & shipping association <http://www.citrus.go.kr>
- Jin SB, Yun SH, Park JH, Park SM, Koh SW, Lee DH (2015) Early identification of citrus zygotic seedlings using pollen-specific molecular markers. *Korean J Hortic Sci Technol* 33:598-604 (in Korean). doi:10.7235/hort.2015.14200
- Kedar VP, Gopal N (1977) Fruitfulness in Nagpur Sangtra (*Citrus reticulata*) as affected by various modes of pollination. *Ind J Hortic* 34:385-386
- Matsumoto R (2001) 'Shiranuhi' a late-maturing citrus cultivar. *Bull Natl Inst Fruit Tree Sci* 35:115-220
- Mondal B, Saha R (2013) Identification of zygotic and nucellar seedlings of *Citrus reticulata* and *Citrus aurantifolia* using RAPD. *Int J Adv Biotechnol Res* 5:25-30
- Moshkov IE, Novikova GV, Hall MA, George EF (2008) Plant growth regulators III: Gibberellins, ethylene, abscisic acid, their analogues and inhibitors; miscellaneous compounds. In EF George, MA Hall, GJ de Klerk, eds, *Plant Propagation by Tissue Culture Ed 3*. Springer, The Netherlands, pp 227-281
- Murashig T, Tucker PH (1969) Growth factor requirements of citrus tissue culture. *Proc 1st Int, Citrus Symp* 3:1155-1161
- Murashige T, Skoog F (1962) A revised medium for rapid growth and bioassays with tobacco tissue cultures. *Physiol Plant* 15:473-497. doi:10.1111/j.1399-3054.1962.tb08052.x
- Ohta Y, Furusato K (1957) Embryo culture in *Citrus*. *Seiken Ziho, Rep Kihara Inst Biol Res* 23:49-54
- Oliveira ACD, Garcia AN, Cristofani M, Machado MA (2002) Identification of *Citrus* hybrids through the combination of leaf apex

- morphology and SSR markers. *Euphytica* 128:397-403. doi:10.1023/A:1021223309212
- Pérez-Tornero O, Porras I** (2008) Assessment of polyembryony in lemon: Rescue and in vitro culture of immature embryos. *Plant Cell Organ Cult* 93:173-180. doi:10.1007/s11240-008-9358-0
- Pieringer AP, Edwards GJ** (1967) Identification of nucellar and zygotic *Citrus* seedlings by infrared spectroscopy. *J Am Soc Hortic Sci* 86:226-234
- Plackett ARG, Wilson ZA** (2016) Gibberellins and plant reproduction. *Ann Plant Rev* 49:323-358. doi:10.1002/9781119210436.ch11
- Rangan TS, Murashige T, Bitters WP** (1969) In vitro studies of zygotic and nucellar embryogenesis in *Citrus*. *Proc 1st Int Citrus Symp* 1:225-229
- Rodriguez M, Monter A, Espinosa A, Castañeda CG, Velázquez A** (2005) Polyembryony and RAPD markers for identification of zygotic and nucellar seedlings in *Citrus*. *Agrociencia* 39:371-383. doi:10.1590/S0100-204X2004000600006
- Ruiz C, Breto MP, Asins MJ** (2000) A quick methodology to identify sexual seedlings in citrus breeding programs using SSR markers. *Euphytica* 112:89-94. doi:10.1023/A:1003992719598
- Sagee O, Shaked A, Hasdai D** (1990) Rooting of cuttings from gibberellin and benzyladenine treated citrus trees. *J Hortic Sci* 65:473-478. doi:10.1080/00221589.1990.11516081
- Schuelke M** (2000) An economic method for the fluorescent labeling of PCR fragments. *Nat Biotechnol* 18:233-234. doi:10.1038/72708
- Sharma RR, Goswami AM, Saxena SK, Shukla A** (1999) Influence of rootstocks on fruit drop in Kinnow mandarin under dense planting. *J Appl Hortic* 1:133-134
- Soost RK, Cameron JW** (1975) *Citrus*. In J Janick, JN Moore, eds, *Advances in Fruit Breeding*. Purdue University Press, West Lafayette, IN, USA, pp 507-540
- Tan ML, Song JK, Deng XX** (2007) Production of two mandarin × trifoliolate orange hybrid populations via embryo rescue with verification by SSR analysis. *Euphytica* 157:155-160. doi:10.1007/s10681-007-9407-5
- Tatum JH, Berry RE, Hearn CJ** (1974) Characterization of *Citrus* cultivars and separating of nucellar and zygotic seedlings by thin layer chromatography. *Proc Florida State Hortic Soc* 87:75-81
- Usman M, Ramzan M, Fatima B, Jaskani MJ, Khan MM** (2002) Citrus germplasm enhancement by interploidy hybridization. 1. Reciprocal crosses of Kinow and Succari. *Int J Agric Bio* 4:208-210
- Woo J, Park YC, Lee JW, Yun S, Kim M, Park S, Lee Y, Song KJ, Kim HB** (2019) Evaluation of polyembryony for genetic resources and efficacy of simple sequence repeat markers for the identification of nucellar and zygotic embryo-derived individuals in citrus. *Appl Biol Chem* 62:30. doi:10.1186/s13765-019-0437-1
- Yelenosky G** (1985) Environmental factors affecting *citrus*. *Fruit Varieties J* 39:51-57
- Yildiz E, Kaplankiran M, Demirköser TH, Uzun A, Toplu C** (2013) Identification of zygotic and nucellar individuals produced from several citrus crosses using SSRs Markers. *Not Bot Horti Agrobot* 41:478-484. doi:10.15835/nbha4129037
- Yun JU, Yang HB, Jung YH, Yun SH, Kim KS, Kim CS, Song KJ** (2007) Identification of zygotic and nucellar mandarin seedling using randomly amplified polymorphic DNA. *Hortic Environ Biotechnol* 48:171-175